

## Interection of Silicon on Heavymetal and Other Stresses in Crop Plants

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### ABSTRACT

*Silicon is the most abundant element in soil and is beneficial for a large variety of plants. It is concentrated in plant tissues in quantities similar to that of macronutrients. Considerable damages to plants caused by abiotic stresses such as drought stress, salinity stress, heavy metal stress and nutrient imbalance, as well as biotic stresses like insect pests and pathogens and even herbivorous attacks, have been reported to be reduced significantly by silicon application. Soil contamination with toxic heavy metals (such as Cd, Pb, As, Hg, Zn) is becoming a most devastating problem worldwide because of the rapid development of social economy. Silicon significantly improved the growth and biomass of crop plants and reduced the toxic effects of heavy metals after different stress periods. Si treatment ameliorated root function and structure compared with non-treated crop plants, which suffered severe root damage. Silicon plays a substantial role in alleviating heavy metal toxicity in crop plants. Also, silicon may reduce the toxic effects of heavy metals in soil. It may protect the foliage and increase light uptake and reduce respiration. Therefore, in this review, we discussed the effects of silicon on heavy metal stress in especially field crops.*

**Key words:** Silicon, Heavy Metal, Field crops, Soil.

### INTRODUCTION

Silicon constitutes a major substantial percentage of different soil types, generally about 31%<sup>31</sup>. In soil solutions, silicon is found mostly as uncharged monomeric silicic acid at concentrations from about 0.1 mM to 0.6 mM<sup>7</sup> or up to about 0.8 mM at equilibrium<sup>20</sup> when the solution pH is below 9<sup>22</sup>. Silicon (Si) is an abundant element in the Earth's crust and plays a role in heavy metal alleviation in plants by different mechanisms<sup>10</sup>. Si reduces the translocation of Cd from roots to shoots and

thus, prevents the adverse effect of Cd on photosynthetic machinery and grains<sup>9</sup>. However, Cd in high concentration is also trapped in roots through vacuolar sequestrations, leading to decreased Cd translocation in aerial parts of the plants. Phytochelatins (PCs) and metallothioneins (MTs) may bind to Cd before transporting the complexes into the vacuole out of the cell by ATP-binding cassette transporters in few plants<sup>15</sup>. PCs are formed from glutathione by the induction of *PCS1* gene<sup>29</sup>.

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Further, MTs are involved in detoxifying cytosolic environment of the cell from Cd toxicity<sup>4</sup>. The heavy metal is taken up in to the cell via carriers, such as low- affinity cation transporters and Fe-regulated transporters in plants<sup>32</sup>. Among the Fe transporters, *IRTs* and *NRAMPs* have been reported to take up heavy metals. *IRT1* is essential for root Fe uptake in response to Fe deficiency but it also accepts Cd as a substrate and is involved in the root-to- shoot transport of Cd<sup>26</sup>. In a transgenic study, elimination of *NRAMP5* transporter reduces Cd uptake in rice<sup>13</sup>. Additionally, the ferric chelate reductase (*FRO*) gene may perform key functions in Fe acquisition in plants. It was reported the inhibition of Fe translocation when bean plants were exposed to chromium (Cr) in nutrient solutions<sup>2</sup>. Also, Cr affects Fe uptake in dicots either by inhibiting the reduction of Fe (III) to Fe (II) or by competing with Fe (II) at the site of absorption. In addition, *IRT1* is induced in response to Fe-deficiency and is capable of transporting minerals and heavy metals<sup>34</sup>. Further, organic acids such as citrate and malate are major chelators in both Strategy I and II plants, which bind Fe at the site of uptake and facilitate long-distance transport in plants<sup>16</sup>. To complete a life cycle, plants are continuously exposed to various abiotic stresses and sometime multiple stresses. These stresses in turn causing the generation of various reactive oxygen species (ROS), such as singlet oxygen ( $1O_2$ ), superoxide ( $O_2^-$ ), hydrogen peroxide ( $H_2O_2$ ), or hydroxyl radicals (OH) in cells. These ROS can cause serious oxidative damage to the protein, DNA, and lipids of cell components<sup>33</sup>. Therefore, ROS scavenging is most important defense mechanism to cope with stress condition in plants<sup>5</sup>. According to previous reports, exogenously Si can improve the ability of ROS scavenging by regulation of antioxidants enzyme activity. Furthermore, regulation pattern across various crop plants is different depending upon the exposure time of the stress<sup>17</sup>. Therefore, here, we discussed various possibilities based on previous literature

survey and our understanding the role of Si in modulating antioxidant activities in plants during abiotic stress.

### DEFENSE MECHANISM AGAINST ROS GENERATION

Plants continuously produce several ROS during metabolic process like photosynthesis and respiration processes in cell organelles such as mitochondria, chloroplast, and peroxisomes. In plants, superoxide dismutase (SOD), catalase (CAT), and ascorbate peroxidase (APX) are the main enzymatic antioxidants, whereas carotenoids, tocopherols, ascorbate, and glutathione are classified as the non-enzymatic antioxidants. SOD are distributed in a different form in various plant organs such as chloroplasts (Cu/ZnSOD, FeSOD), cytosol (Cu/ZnSOD), and mitochondria (MnSOD). Primarily, SOD catalyzes the efficient removal of superoxide free radicals in chloroplasts as they are mainly generated in the photosystem I, during the light reaction. CAT is located in the peroxisomes of plant cells, and its main role is the elimination of  $H_2O_2$ , which is produced by the SOD reaction. Another antioxidant, APX, also can remove  $H_2O_2$ ; however, it is distributed in the peroxisomes as well as chloroplasts, cytosol, and mitochondrion. Plants can induce defense responses against oxidative stress by activating the non-enzymatic antioxidants, which represent the second line of defense against ROS, hydrophilic molecules (ascorbate, glutathione), and lipophilic metabolites (carotenoids,  $\alpha$ -tocopherol<sup>8</sup>. In addition, glutathione protects the thiol-groups of enzymes located in the chloroplast stroma and participates in the production of  $\alpha$ -tocopherol and ascorbate<sup>24</sup>. Besides its role in detoxification of ROS, glutathione induces physiological responses such as the regulation of sulfur transport and expression of stress defense genes. Carotenoids are a class of phenolic compounds distributed in various fruits and vegetables. They can prevent lipid peroxidation by scavenging single oxide radical from chloroplasts<sup>18</sup>.

**Cadmium Toxicity:**

Cadmium, as a non-essential element, is one of the aggravating factors in soil salinity, which plays a major role in inhibition of plant growth by accumulation in plant. Cadmium in the plant could intervene in plant chemical synthesis processes such as ammonification, nitrification, DE nitrification, and microbiological process that affect the quantity and quantity of the crop products<sup>23</sup>. It also leads to the generation of [e.g., “Reactive Oxygen Species (ROS)”] and oxidative stress so that it can impact on the performance of protein and lipids. Cadmium in leaf leads to leaf chlorosis<sup>11</sup>. Photosynthesis inhibition with the decline of pigment content, chlorophyll a, and phycobiliproteins<sup>14</sup>. Results indicated that salicylic acid and silicon alleviate the inhibitory effects of cadmium on maize seedlings by increasing both their chlorophyll content and fresh weight. Although individual treatments of salicylic acid and silicon reduced plants free proline, soluble sugars and cadmium uptake and lipid peroxidation rate, they improved root and shoot fresh weights in both cadmium stressed and unstressed seedlings. When combined, salicylic acid and silicon alleviated the inhibitory effects of cadmium on seedlings significantly<sup>28</sup>.

**Lead (Pb) Toxicity:**

Lead as a non-redox active metal, by positioning in group 14 of the periodic table and having a low melting point is one of the important metals in a variety of industrial products, including paints, weights, ammunitions, and leaded glass. Lead is considered as an immobilized property in the soil so that plants can easily access it; however, it should be noticed how lead enters the plant body. One of the consequences of increasing lead is the production of ROS in plant cells, which can cause the replacement of essential ions in the cell and impair other processes such as cell adhesion and cell signaling<sup>27</sup>. In the cell, nuclear by binding with DNA, lead can reduce the role of repairs in DNA and lead to a disturbance in mitotic stage and prolongs interface and consequently,

increase the period of the cell cycle<sup>21</sup>. Pb (lead toxicity) in plants can decrease the growth of roots and increase the roots' suberized. Pb (lead), with impact on the Reaction Centre and Antennae, decreases the efficiency of photosystem II, which can negatively affect plant metabolism. The key mechanisms of Si mediated toxicity alleviation mechanisms is mainly 1) Complexation and co-precipitation of toxic metals with silicon, 2) Immobilization of toxic metals ions 3) Compartmentalization with vacuoles. Silicon (Si) addition protect the plant tissues from membrane oxidative damage under Pb stress, thus mitigating Pb toxicity and improving the growth of cotton plants<sup>19</sup>. The results of the present experiment coincided with the conclusion that Silicon (Si) is involved in the metabolic or physiological changes in Pb stressed cotton plants<sup>3</sup>.

**Silicon and other stress interection:**

Silicon can reduce the negative effects of other stresses including physical stresses (high temperature, freezing, drought, lodging, radiation, irradiation, UV) and chemical stresses (salt, nutrient imbalance, metal toxicity) in *Borago officinalis* L. Probably, because of the strengthening effects on cell wall. Silicon has indicated a significant effect on lodging especially in rice, wheat and barley by enhancing the amount of light and photosynthesis<sup>6</sup>. Some researchers reported a highly significant role of supplied silicon in enhancing the biosynthesis of phenolic compounds under UV-B stress. The result of experiments showed that silicon application was significant in alleviating the adverse effects of UV-B. Significant advances have been made in alleviating the effects of UV-B by exogenous silicon application on soybean, wheat and maize<sup>30</sup> silicon application reduced the apoplastic Mn levels in cowpea<sup>12</sup>. The most significant effects of silicon on metal toxicity are by reducing Cd and copper (Cu) uptake and root-to-shoot translocation by increasing metal adsorption and Zn and Mn uptake<sup>25</sup>. It was revealed that silicon reduced Cd uptake by the plants as well as decreased shoot to grain translocation of Cd<sup>1</sup>.

### CONCLUSION

The mechanism responsible for increased metal tolerance in Si-treated plants is still a matter of discussion and contradictory results have been reported. Si application shows varying response to ROS scavenging by activating the defense system plants. In doing so, the activity of antioxidant (CAT, SOD, PPO, POD, APX, GPX, and GSSH) may also oscillate depending upon the intensity of heavy metal stress and plant type. However, plants treated with Si presented not only biomass increasing but also higher metal accumulation. This clearly indicates that a silicon-mediated mechanism plays a role in alleviating the metal stress. Significant structural alterations on xylem diameter, mesophyll and epidermis thickness, and transversal area occupied by collenchyma and midvein were also observed as a result of Si application. The precipitation of silica in the endodermis and pericycle of roots seems to play an important role on the crop plants tolerance to heavy metal stress. Such results indicate that Si could be used in phytotechnologies aiming at increasing the tolerance and accumulation of metals in plants, which may become very good opportunity to reduce heavy metals from our food grains.

### REFERENCES

1. Adrees, M., Ali, S., Rizwan, M., Rehman, M.Z., Ibrahim, M., Abbas, F., Farid, M., Qayyum, M.F., Irshad, M.K., Mechanisms of silicon-mediated alleviation of heavy metal toxicity in plants: A review. *Ecotoxicology and Environmental Safety*, **119**: 186-197 (2015).
2. Barcelo, J., Poschenrieder, C., Vazquez, M. D., Gunse, B., and Vernet, J. P., Beneficial and toxic effects of chromium in plants: solution culture, pot and field studies. *Studies in Environmental Science No. 55*: Paper Presented at the 5th International Conference on Environmental Contamination, Morges (1993).
3. Bharwana SA , Ali S., Farooq MA , Iqbal N , Abbas F and Ahmad MSA., Alleviation of Lead Toxicity by Silicon is Related to Elevated Photosynthesis, Antioxidant Enzymes Suppressed Lead Uptake and Oxidative Stress in Cotton. *J Bioremed Biodeg* , **4**: 4 (2013).
4. DalCorso, G., Farinati, S., and Furini, A. Regulatory networks of cadmium stress in plants. *Plant Signal. Behav.* **5**: 663–667(2010).
5. Das, K., and Roychoudhury, A. Reactive oxygen species (ROS) and response of antioxidants as ROS-scavengers during environmental stress in plants. *Front Environ Sci.* **2**: 53 (2014).
6. Dorairaj, D., Ismail, M.R., Sinniah, U.R., Ban, T.K., Influence of silicon on growth, yield, and lodging resistance of MR219, a lowland rice of Malaysia. *Journal of Plant Nutrition*, **40**: 8 (2017).
7. Epstein E . The anomaly of silicon in plant biology. *Proceedings of the National Academy of Sciences of the United States of America* **91**: 11–17 (1994).
8. Gowayed, S. M., Al-Zahrani, H. S., and Metwali, E. M. Improving the Salinity tolerance in potato (*Solanum tuberosum*) by exogenous application of silicon dioxide nanoparticles. *Int. J. Agric. Biol.* **19**: 183–194.(2014).
9. Greger, M., and Landberg, T. “Influence of silicon on cadmium in wheat,” in Proceedings of the 4th International Conference on Silicon in Agriculture, ed. M. Laing (Durban: Aim Print), 25 (2008).
10. Greger, M., Kabir, A. H., Landberg, T., Maity, P. J., and Lindberg, S. Silicate reduces cadmium uptake into cells of wheat. *Environ. Pollut.* **211**: 90–979 (2016).
11. H .Lin, C .Fang, Y .Li, W .Lin, J .He, R .Lin, and W. Lin. "Effect of silicon on grain yield of rice under cadmium-stress", *Acta Physiol Plant* ,vol.**38**,pp.186 (2016).
12. Horst, W.J., Fecht, M., Naumann, A., Wissemeier, A.H., Maier, P. Physiology of manganese toxicity and tolerance in *Vigna unguiculata* (L.) Walp. *Journal of Plant Nutrition and Soil Science*, **162**: 263-274 (2016).

13. Ishikawa, S., Ishimaru, Y., Igura, M., Kuramata, M., Abe, T., Senoura, T., et al. Ion-beam irradiation, gene identification, and marker-assisted breeding in the development of low-cadmium rice. *Proc. Natl. Acad. Sci. U.S.A.* 109, 19166–19171(2008).
14. Simek, J. Tuma, V. Dohnal, K. Musil, and Z. Ducaiova. "Salicylic acid and phenolic compounds under cadmium stress in cucumber plants (*Cucumis sativus* L.)" *Acta Physiol Plant*, **38**: pp.172 (2016).
15. Jasinski, M., Ducos, E., Martinoia, E., and Bounry, M. The ATP-binding cassette transporters: structure, function, and gene family comparison between rice and *Arabidopsis*. *Plant Physiol.* **131**: 1169–1177 (2003).
16. Kabir, A. H., Paltridge, N. G., Able, A. J., Paull, J. G., and Stangoulis, J. C. R. Natural variation for Fe-efficiency is associated with upregulation of Strategy I mechanisms and enhanced citrate and ethylene synthesis in *Pisum sativum* L. *Planta* **235**: 1409–1419 (2012).
17. Kim, Y. H., Khan, A. L., Waqas, M., Shahzad, R., and Lee, I. J. Silicon-mediated mitigation of wounding stress acts by up-regulating the rice antioxidant system. *Cereal Res. Commun.* **44**: 111–121 (2016).
18. Kühlbrandt, W., Wang, D. N., and Fujiyoshi, Y. Atomic model of plant light-harvesting complex by electron crystallography. *Nature* **367**: 614–621 (1994).
19. L. H. D. Dao, and J. Beardall, "Effects of lead on two green microalgae *Chlorella* and *Scenedesmus*: photosystem II activity and heterogeneity." *Algal Research*, **16**: pp.150–159 (2016).
20. Lindsay WL. Chemical equilibria in soils, John Wiley and Sons, New York. 51–54 (1979).
21. Liu, J., Ma, J., He, C., Li, X., Zhang, W., Xu, F., et al. Inhibition of cadmium ion uptake in rice (*Oryza sativa*) cells by a wall-bound form of silicon. *New Phytol.* **200**: 691–699 (2013).
22. M. Dikilitas, S. Karakas, and P. Ahmad, "Effect of lead on plant and human DNA damages and its impact on the environment", *Plant Metal Interaction* (2016).
23. Ma JF, Takahashi E. Soil, fertilizer and plant silicon research in Japan, 1st ed. *Elsevier*, Amsterdam.(2002)
24. P. Cojocar, Z.M. Gusiatin, and I. Cretescu, "Phytoextraction of Cd and Zn as single or mixed pollutants from soil by rape (*Brassica napus*)", *Environ Sci Pollut Res Int*, **23(11)**: pp.10693-701 (2016).
25. Racchi, M. L. Antioxidant defenses in plants with attention to *Prunus* and *Citrus* spp. *Antioxidants* **2**: 340–369 (2013).
26. Rizwan, M. Silicon-mediated heavy-metal tolerance in durum wheat: Evidences of combined effects at the plant and soil levels. These pour obtenir le grade de Docteur d'Aix-Marseille universite, Faculté des Sciences et Techniques, *Discipline: Géosciences de l'Environnement, France.* ([www.theses.fr/2012AIXM4335/abes](http://www.theses.fr/2012AIXM4335/abes)) (Date of Access: **20**: 03.2017).
27. Rogers, E. E., Eide, D. J., and Guerinot, M. L. Altered selectivity in an *Arabidopsis* metal transporter. *Proc. Natl. Acad. Sci. U.S.A.* **97**: 12356–12360 (2000).
28. S. Lyer, C. Sengupta, and A. Velumani, "Lead toxicity: An overview of prevalence in Indians", *Clin. Chim. Acta*, **451**: 161–164 (2015).
29. S. Mohsenzadeh1, M. Shahrtash1 and H. Mohabatkar1, 2011. Interactive effects of salicylic acid and silicon on some physiological responses of cadmium-stressed maize seedlings. *Iranian Journal of Science & Technology, IJST A1*: 57-60 (2011).
30. Semane, B., Cuypers, A., Smeets, K., Van Bellegem, F., Horemans, N., Schat, H., et al. Cadmium responses in *Arabidopsis thaliana*: glutathione metabolism and antioxidative defence system. *Physiol. Plant.* **129**: 519–528 (2007).

31. Shen, X., Li, Z., Duan, L., Eneji, A.E., Li, J. Silicon mitigates ultraviolet-B radiation stress on soybean by enhancing chlorophyll and photosynthesis and reducing transpiration. *Journal of Plant Nutrition*, **37(6)**: 837-849 (2014).
32. Sposito, G., The chemistry of soils. Oxford University Press, New York (1989).
33. Takahashi, R., Ishimaru, Y., Senoura, T., Shimo, H., Ishikawa, S., Arao, T., The Os NRAMP1 iron transporter is involved in Cd accumulation in rice. *J. Exp. Bot.* **62**: 4843–4850 (2011).
34. Tripathi, D. K., Singh, S., Singh, V. P., Prasad, S. M., Dubey, N. K., and Chauhan, D. K. Silicon nanoparticles more effectively alleviated UV-B stress than silicon in wheat (*Triticum aestivum*) seedlings. *Plant Physiol. Biochem.* **110**: 70–81 (2017).
35. Vert, G., Grotz, N., Dedaldechamp, F., Gaymard, F., Guerinot, M. L., Briat, J., IRT1, an Arabidopsis transporter essential for iron uptake from the soil and for plant growth. *Plant Cell* **14**: 1223–1233 (2002).